OneArray®

Rice OneArray® User Guide

www.OneArray.com

Phalanx Biotech Group

Notice to the User

It is important that users read the entire manual before commencing work.

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Contact Information

✧ Asia Office

Phalanx Biotech Group
6 Technology Road 5, 6th Floor
Hsinchu Science Park
Hsinchu 30078, Taiwan

Ph: 886.3. 5781168
Toll-Free: 0800-777-988
FAX: 886.3. 5785099
E-mail: twsales@phalanxbiotech.com
website: www.onearray.com

✧ U.S. Office

Phalanx Biotech Group
1301 Shoreway Road, Suite 160
Belmont, CA, USA, 94002

Ph: 1.650.320.8669
Toll-Free: 877.730.3887
FAX: 1. 650.508.9889
E-mail: info@phalanxbiotech.com
website: www.onearray.com

✧ User Guide and Technical Support

Electronic version of this manual is available on the enclosed Product Support CD, and online at:
www.onearray.com

To reach technical support by telephone, call
Within the US: 1.650.320.8669
Outside the US: 886.3.5781168

✧ Feedback

We welcome your feedback regarding our products and this manual. Please contact us at:
twsales@phalanxbiotech.com

All comments are welcome.

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Thank You

Phalanx Biotech Group would like to extend special thanks to our customers who have provided feedback that enabled us to improve the OneArray® User Guide.

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Getting Started

Please read the introductory information below to help familiarize yourself with OneArray® before use.

Product Contents

- Rice OneArray® DNA Microarray
- OneArray® Hybridization Buffer Tube

- Each tube contains buffers sufficient for 50 microarray hybridization procedures
- Spare round cap tube
- OneArray® User Guide
- Spotted Region Guide
- Product Support CD, which contains the following:

- Sample Images
- OneArray.gal file
- OneArray® gene list and probe sequences
- OneArray® microarray layout
- OneArray® Control Probe list
- OneArray® User Guide (electronic version)

Other Necessary Apparatus (Not Supplied)

Apparatus

- Water bath/heating block
- Powder-free gloves
- Clean, blunt forceps
- Micropipettors
- Sterilized and nuclease-free pipet tips
- Sterilized and nuclease-free microcentrifuge tubes
- High-speed microcentrifuge
- Low-speed tabletop microcentrifuge with slide holder attachment
- Vortex mixer
- Hybridization oven
- Hybridization accessories: chamber cover slides, etc.
- Rectangular slide staining dish and slide rack for washing microarrays
- PCR (polymerase chain reaction) machine
- Microarray scanner for standard 1" x 3" format (see Table 8 under "OneArray® Microarray Scanner Specifications" for a list of compatible scanners)
- Hybridization systems (optional)
- Automated hybridization station (optional)

Other Necessary Reagents (Not Supplied)

Reagents

- De-ionized nuclease-free **water**
- Cyanine 3- or 5-labeled amplified **aRNA** sample
- **20X SSC stock** solution, sterile filtered:
  - 3.0 M Sodium chloride
  - 0.3 M Sodium phosphate (pH 7.0)
- **20X SSPE stock** solution, sterile filtered:
  - 3.6 M Sodium chloride
  - 0.2 M Sodium phosphate (pH 7.7)
  - 20 mM EDTA
- **Wash Solutions**, sterile filtered (four types, approximately 250 mL of each is required per experiment):
  - 2 X SSC, 0.2% SDS
  - 2 X SSC
  - 0.2 X SSC

  NOTE: ***SDS must be molecular biology grade.***
- **Pre-hybridization Buffer**, prepared and sterile filtered immediately prior to pre-hybridization:
  - 5X SSPE, 0.1% SDS, 1% BSA

  NOTE: ***BSA must be molecular biology grade.***
- Deionized **formamide** to be added to the OneArray® Hybridization Buffer prior to use (see Step 4).
- RNA **Fragmentation Reagent and Stop Solution** (for hybridization using aRNA)
- **DNA** Blocking Mixture:

- Ambion® sheared Salmon Sperm DNA (10 μg/μl), or Invitrogen™ Cot-1 DNA® (2.5 10 μg/μl), or Invitrogen™ Poly-A (2.5 10 μg/μl)

Important Notes on Microarray Handling and Storage

Storage Conditions

- Store unopened OneArray® product at room temperature.
- Store opened OneArray® product at room temperature.
- Store OneArray® Hybridization Buffer at room temperature.

NOTE: ***If the product is received with an open bag, please contact Phalanx Biotech Customer Service for an immediate replacement.***

Handling Microarrays

Please read this section carefully and follow the instructions!

- Polynucleotide probes are printed on the side of the slide with the barcode.
- To avoid irreparable damage of the printing area, **do not touch** the surface with bare hands, or with any other objects.
- Whenever possible, handle microarrays with clean blunt forceps to avoid contamination.

Open arrays should be used within a week.

Product Descriptions and Overview

OneArray® Whole Genome DNA microarrays are made of sense-strand polynucleotide probes spotted onto a proprietary chemical layer coated on top of a 1" x 3" (25 mm x 75 mm) standard-format microarray glass slide. Updated information of genome content from public domains is used to design approximately 22,000 highly sensitive long-oligonucleotide probes for monitoring the expression level of corresponding protein-coding genes.

Each probe is spotted onto the array in a highly consistent manner using a proprietary, non-contact spotting technology adapted for microarray manufacturing.

Rice OneArray® v1 Genome Content

Each microarray contains 22,003 oligonucleotides including 21,179 rice genome probes, and 824 experimental control probes. Based on "One Gene-One Probe" design concept, probe is designed to hybridize to a specific target gene described in the current public domain contents, such as RGAP (Rice Genome Annotation Project) database v6.1 and BGI (Beijing Genomics Institute) database release in 2008.

Table 1, below, provides an example of the contents of a Rice genome that can be studied using the Rice OneArray® v1.

Table 1: Rice* OneArray® v1 probe *Content

Probe Type	Number of Probes
RiOA 1.0 probes	**22,003 (total)***
New Probes Design based on: **- RGAP v6.1** **- BGI 2008**	**21,179**
Control Probes	**824**

* Rice OneArray® v1 is guaranteed to contain > 98% of the total probe content.

Rice OneArray® v1 Control Features

There are 824 control probes built into the Rice OneArray® DNA microarray that monitor the sample quality and hybridization process. These control probes provide valuable information to ensure experiments are done correctly to ensure higher quality results for analysis.

NOTE: ***Detailed control information, gene lists, gene annotations, and probe sequences can be found on the Product Support CD that accompanied this product, or at: http://www.onearray.com***

Using OneArray®

This section provides you with detailed information about how to perform the steps necessary to complete the hybridization process to study gene expressions using the OneArray® microarray.

Follow these detailed steps *exactly* to achieve the best experimentation results.

- **Step 1:** Prepare the RNA Sample
- **Step 2:** Label the Target
- **Step 3:** Pre-Hybridize the Microarray
- **Step 4:** Perform the Hybridization Protocol
- **Step 5:** Wash the Hybridized Microarray
- **Step 6:** Scan and Extract Gene Expression Results
- **Step 7:** Check Control Probe Data

Step 1: Prepare the RNA Sample

High-quality, intact RNA is essential for all gene expression microarray experiments.

There are many different RNA isolation protocols and commercially available RNA isolation kits. You should choose a solution that meets your specific needs. Qiagen, Ambion, Invitrogen and other reagent companies offer many different RNA isolation products. For more information, you can visit each company's website.

Once the RNA samples are isolated, you must confirm the quantity and quality of the samples. Similarly, many different protocols are available and you should choose a solution that is suitable for your needs.

For faster and more automated RNA analysis, you may want to consider the "No Cuvettes" Spectrophotometer from NanoDrop™, or the 2100 Bioanalyzer from Agilent Technologies. For more information, visit each company's website.

Step 2: Label the Target

For best results, it is recommended that you use one of the commercially available labeling kits that has been tested for use with the OneArray® microarray—please refer to Tables 3 and 4 below.

General Guidelines for Target Labeling

There are many commercially available labeling kits for microarray analysis. Select a labeling kit or labeling method that is most suitable for your specific needs. If you use a labeling kit that is not listed in Tables 3, it is recommended that you validate the method to test and determine its compatibility with the OneArray® .

You may want to confirm the quality of the labeled target with the "No Cuvettes" Spectrophotometer from NanoDrop™.

RNA Sample Amounts

Generally, the amount needed of quality RNA is 10 μg for each labeling reaction.

If you have an *ample* supply of RNA samples, you have the *choice* of using a protocol that either amplifies or does not amplify the RNA sample.

If you have a *limited* amount of RNA samples, it is recommended that you use a protocol that includes a linear amplification of the RNA samples.

Dye Incorporation Efficiency

Good dye incorporation rates are important for yielding the best data from microarray hybridization. Incorporation rates of 20-60 dye molecules per 1,000 bases (20-33 bases / dye molecule) yield the most usable data. Rates below 20 dyes per 1,000 bases (50 bases / dye) are very low and may lead to a loss of signal of many targets. It is not recommended to perform hybridization with samples of low dye incorporation efficiency.

For aRNA Hybridization

Follow the instructions provided by the reagent supplier. Indirect labeling with NHS ester dye is recommended. Table 3, below, contains a list of products that have been tested for use with OneArray®.

Table 3: aRNA Preparation Products

Manufacturer	Product Name and Description
Ambion®	Amino Allyl MessageAmp II™ aRNA Kit
Ambion®	aRNA Fragmentation Reagent
Epicentre® Biotechnologies	TargetAmp™ 1-Round Aminoallyl - aRNA Amplification Kit

For aRNA labeling, 10 µg of quality aRNA is recommended. Smaller volumes can lead to significant loss of sample and may increase the concentration of contaminants in the labeled aRNA sample, leading to higher background signal.

It is best to use aRNA as soon as possible after labeling, as exposure to air and light can reduce the signal of some dyes. If it must be left overnight, it is best to aliquot your labeled aRNA and store in the dark at -80°C. Avoid thawing and re-freezing aRNA if possible, as freeze-thaw cycles can damage the aRNA.

Finally, aRNA fragmentation is best performed immediately prior to hybridization (Step 4).

Step 3: Pre-Hybridize the Microarray

General Instructions

OneArray® requires a pre-hybridization step prior to hybridization of the labeled target. The pre-hybridization step reduces background signals and increases the performance of the microarray. Complete the pre-hybridization step by carefully following the instructions below.

1) Warm the pre-hybridization solution (5X SSPE, 0.1% SDS, and 1% BSA) to 42 °C.

2) Pour 25 ml room temperature 100% ethanol into the spare array tube.

3) Preheat the OneArray® (s) in the round cap tube at 60 °C for 10 min (hybridization oven recommended).

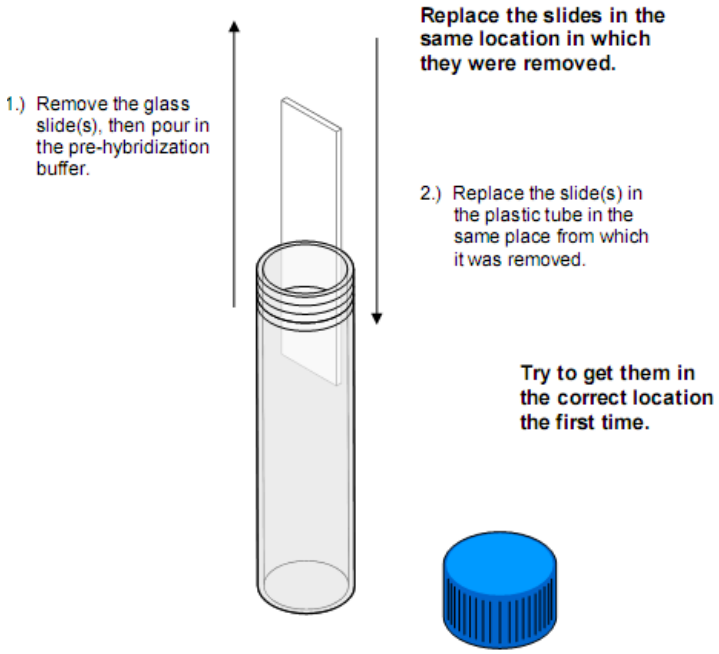
4) Remove the OneArray® (s) from the round cap tube, place in the two outermost slots inside the tube containing 100 % ethanol, close the cap, and let sit for approximately 15 sec.

5) Shake the round cap tube for 20 sec.

6) Remove and thoroughly rinse each array with deionized water to remove any residual ethanol.

7) Carefully and slowly, fully submerge the OneArray® in an abundant amount of pre-hybridization solution for 1 hr at 42 °C (35 ml is sufficient if using a round cap tube).

Try to insert the slides into the correct position the first time. Avoid inserting and removing the slides more than once in the pre-hybridization buffer.


Replace the slides in the same location in which they were removed.
1.) Remove the glass slide(s), then pour in the pre-hybridization buffer.
2.) Replace the slide(s) in the plastic tube in the same place from which it was removed.
Try to get them in the correct location the first time.

8) After 1 hr, transfer the slide(s) to room temperature, distilled water and wash gently for 2 min.

9) Spin dry the slide(s) for 2 min. Store in a dry, dark place until hybridization. It is recommended that you use the slides in the hybridization protocol within 1 hr of completing the pre-hybridization process.

Step 4: Complete the Hybridization Protocol

Once you have completed the pre-hybridization step using one of the methods outlined in the Step 3: Pre-Hybridize the Microarray section, you are ready to complete the hybridization protocol.

There are many different hybridization protocols, apparatus, and instruments available that may be compatible for use with the OneArray® microarray. Detailed instructions for using Phalanx hybridization system, OneArray® Full Length Chamber and the glass cover slide method are described below.

For best performance and consistent hybridization results, it is recommended that you use the OneArray® Hybridization Buffer, included with this product to complete the hybridization process.

A. Using the Phalanx hybridization system

Step 4Aa: →Prepare Hybridization Solution Using the OneArray® Hybridization Buffer (Included)

For correct use of this buffer, you must add a specific amount of formamide and labeled target. Please follow the instructions below carefully.

1) Spin down the stock OneArray® Hybridization Buffer (~ 5.5 ml in each tube).

2) Add 4.5 ml of deionized formamide.

3) Warm the mixture to 42 °C to completely dissolve the solution. Mix thoroughly.
   Yield: 10 ml of 1.5X Hybridization Buffer solution.

4) Make up 1 X Hybridization Buffer by adding nuclease-free H_2O.

5) Aliquot the solution into individual tubes according to usage and store in darkness at -20 °C.

Step 4Ab: →Prepare Target for Hybridization

✧Hybridization Using Labeled Targets from aRNA Labeling Approaches

1) Mix 10 µg of your aRNA sample with nuclease-free H_2O to yield a final volume of 27 µL.

NOTE: ***It is essential to use at least 10 µg of labeled target for each hybridization. If you are performing a dual-dye experiment, use at least 10 µg of each labeled aRNA sample.***

2) Add 3 µl 10x Fragmentation Reagent, and incubate at 70 °C for 15 minutes.

3) Add 3 µl Stop Solution, and mix well.

4) Mix with nuclease-free H_2O to yield a final volume of 60 µL.

5) Keep on ice and in darkness until hybridization (Step 4Ac).

Step 4Ac: →Complete the Hybridization

1) Thaw and re-suspend the 1.5X and 1X Working Hybridization buffer at 42~65 °C for 10 minutes.

2) Preheat water bath to 95 °C. The water batch should be sufficient to submerge a microarray slide vertically, i.e. large beaker.

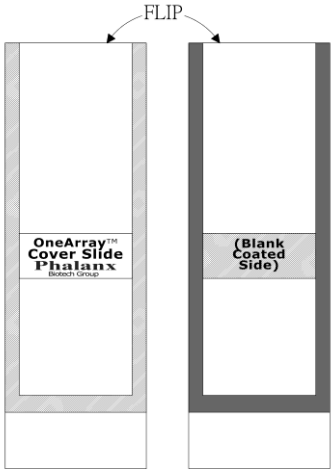
3) Prepare Target Hybridization Mix:

Final Total Volume of Target Hybridization Mix	180 µl
Labeled target mix	**60 µl**
1.5X Working Hybridization Buffer **Add RNAase free ddH_2O to reach the final volume**	**120 µl**

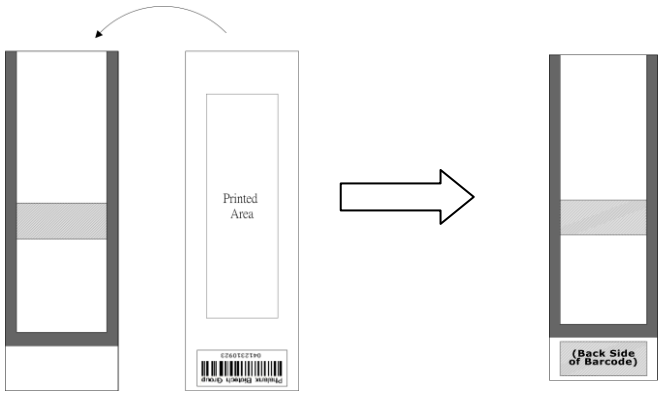
NOTE: ***Different volumes of labeled target mix may be obtained due to different labeling protocols. If the final volume of the labeling target mix is less, use distilled water to make up the volume.***

4) Denature the Target Hybridization Mix from the previous step in a PCR machine at 95 ^{o}C for 5 minutes and hold at 60 ^{o}C.

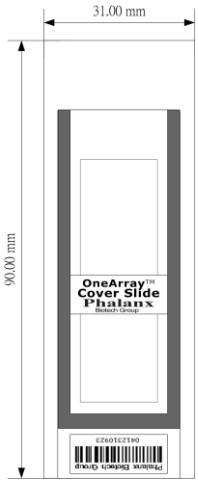
5) Assembling Process:



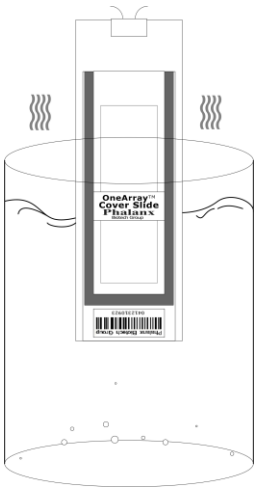
i. Take cover slide out from the package.



ii. Carefully laid the Rice OneArray® on top of the assembly of cover slide and spacer, where the printed side of array should be facing toward the cover slide, to form a hybridization assembly. The printed side of array is on the side of the label



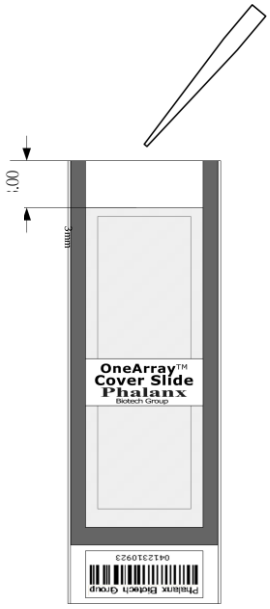
iii. Insert the hybridization assembly into a (3.1 x 9cm) heat-shrank hybridization (Hyb.) bag. Sample loading side of the hybridization assembly should face the opening side of film bag. Lower the assembly to the end of the bag (Assembly viewed from the other side).



iv. Clip the bag with a clipper "securely" from the opening end. Immerse the assembly to the rim of the slide in 95 °C hot water swiftly (approximately 2~5 sec). Do not dip the assembly too far into the water to avoid water leaking into the assembly. The Hyb. bag will shrink and tightly wrap around the assembly.

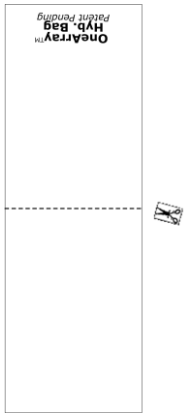
Caution: Hot water bath is >90℃! Do not dip your hand into the water!

v. Remove the assembly from water and wipe off the water from the assembly. Trim off the excess film from the top of the assembly. Keep the assembly in a 50°C oven for at least 10 minutes

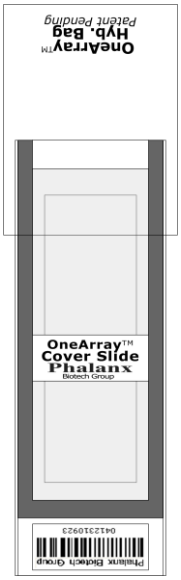


vi. Load the entire target hybridization mix into the assembly through the sample loading opening. Add 60ul of 1X hybridization buffer and leave approximately 3 mm (0.3cm) of space from the top rim.

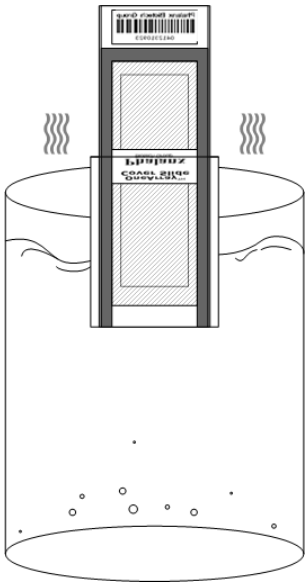
Caution: Tilt the assembly slightly for easier loading.



vii. Cut the second Hyb. Bag to approximately one-half of its original length.



viii. Keep the assembly vertical and slip on the shortened Hyb. bag from step G all the way until the assembly reaches the end of the Hyb. bag.



ix. Use a pair of forceps to hold onto the top of the assembly. Immerse the entire assembly into 95℃ hot water with the assembly in upright position.

Caution: Make sure a bubble forms in the closed hybridization system. If the bubble does not appear or appears to be much smaller, please try to tap on the slide slightly to disturb the mixture into forming a bubble. Also, make certain that all the air bubbles in the mixture eventually mix into one big bubble.

x. Place the hybridization assembly into a 50 °C oven and set it on a rotator. Remember to rotate the assembly at 2 rotations per minute (RPM) for overnight.

Note: Please check to see that the bubble rotates and move freely in the solution to aid in the mixing effect during hybridization.

B. Using OneArray® Full Length Chamber

Step 4Ba: →Prepare Hybridization Solution Using the OneArray® Hybridization Buffer (Included)

For correct use of this buffer, you must add a specific amount of formamide and labeled target. Please follow the instructions below carefully.

1) Spin down the stock OneArray® Hybridization Buffer (~ 5.5 ml in each tube).

2) Add 4.5 ml of deionized formamide.

3) Warm the mixture to 42 °C to completely dissolve the solution. Mix thoroughly.
   Yield: 10 ml of 1.5X Hybridization Buffer solution.

4) Make up 1 X Hybridization Buffer by adding nuclease-free H_2O.

5) Aliquot the solution into individual tubes according to usage and store in darkness at -20 °C.

Step 4Bb: →Prepare Target for Hybridization

✧Hybridization Using Labeled Targets from aRNA Labeling Approaches

1) Mix 10 µg of your aRNA sample with nuclease-free H_2O to yield a final volume of 27 µL.

NOTE: ***It is essential to use at least 10 µg of labeled target for each hybridization. If you are performing a dual-dye experiment, use at least 10 µg of each labeled aRNA sample.***

2) Add 3 µl 10x Fragmentation Reagent, and incubate at 70 °C for 15 minutes.

3) Add 3 µl Stop Solution, and mix well.

4) Mix with nuclease-free H_2O to yield a final volume of 67 µL.

5) Keep on ice and in darkness until hybridization (Step 4Bc).

Step 4Bc: →Complete the Hybridization

1) Thaw and re-suspend the 1.5X and 1X Working Hybridization buffer at 42~65 °C for 10 minutes.

2) Preheat water bath to 95 °C. The water batch should be sufficient to submerge a microarray slide vertically, i.e. large beaker.

3) Prepare Target Hybridization Mix:

Final Total Volume of Target Hybridization Mix	200 µl
Labeled target mix	67 µl
1.5X Working Hybridization Buffer Add RNAase free ddH_2O to reach the final volume	133 µl

NOTE: ***Different volumes of labeled target mix may be obtained due to different labeling protocols. If the final volume of the labeling target mix is less, use distilled water to make up the volume.***

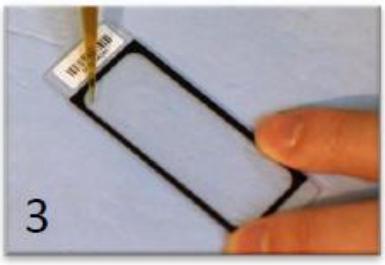
4) Denature the Target Hybridization Mix from the previous step in a PCR machine at 95 ^{o}C for 5 minutes and hold at 60 ^{o}C.

5) Assembling Process:

i. Remove the clear liner on the back of the hybridization chamber. Align the tab-end of the chamber to the edge of the microarray opposite to the barcode. It is easier to hold the long edges of the chamber in one hand and press down the tab with the other hand.(Figure 1)

ii. Use the applicator stick provided to press along the adhesive areas to ensure a secure seal. Visually inspect the seal from underneath the microarray; inconsistent patterns in the black adhesive may indicate an insecure seal. Re-use the applicator stick if needed (Figure 2)

iii. Allow the adhesive to set for 30 minutes



iv. Pipette 200 µL of the labeled RNA solution through one port of the chamber while allowing air to escape through the other port. Make sure there are no bubbles in the pipette tip. If air bubbles form within the chamber, light pressure may be applied to the surface to dislodge them. (Figure 3)

v. Wipe excess solution from the ports. Be careful not to draw solution from the chamber.

vi. Cover ports with supplied circular seals. Seals should be removed from the liner and applied using forceps. The seals will adhere to most wet surfaces. Apply pressure to both seals simultaneously to ensure a secure adhesion. (Figure 4)

vii. Keep the chamber/microarray assembly at 50℃ for 14-16 hrs. Rotation of the assembly during hybridization has been shown to increase the signal intensity

viii. Prepare the first wash solution of 2X SSC, 0.2% SDS and warm to 42℃. More details can be found in the OneArray™ Microarray User Guide.

ix. Remove the chamber/microarray assembly from the hybridization oven and completely submerge it under the wash solution. Use forceps to slowly lift and remove the chamber starting from the tab-end. Use the holes in the tab for a better grip. Be sure to keep the microarray under the wash solution during removal. (Figure 5)

x. Wash the array in the solution, and proceed to follow the remaining steps according to the step 5.

C: Hybridization Using the Glass Cover Slide Method

Step 4Ca: →Prepare Hybridization Solution Using the OneArray® Hybridization Buffer (Included)

For correct use of this buffer, you must add a specific amount of formamide and labeled target. Please follow the instructions below carefully.

1) Spin down the stock OneArray® Hybridization Buffer (~ 5.5 ml in each tube).

2) Add 4.5 ml of deionized formamide.

3) Warm the mixture to 42°C to completely dissolve the solution. Mix thoroughly.
Yield: 10 ml of 1.5X Hybridization Buffer solution.

4) Make up 1 X Hybridization Buffer by adding nuclease-free H_2O.

5) Aliquot the solution into individual tubes according to usage and store in darkness at -20°C.

Step 4Cb: →Prepare Target for Hybridization

✧**Hybridization Using Labeled Targets from aRNA Labeling Approaches**

1) Mix 2 µg of your aRNA sample with nuclease-free H_2O to yield a final volume of 9 µL.

NOTE: ***It is essential to use at least 2 µg of labeled target for each hybridization. If you are performing a dual-dye experiment, use at least 2 µg of each labeled aRNA sample.***

2) Add 1 µl 10x Fragmentation Reagent, and incubate at 70°C for 15 minutes.

3) Add 1 µl Stop Solution, and mix well.

4) Mix with nuclease-free H_2O to yield a final volume of 17 µL.

5) Keep on ice and in darkness until hybridization (Step 4Bc).

Step 4Cc: →Complete the Hybridization

NOTE: ***If you perform hybridization using methods other than the basic glass cover slide method, it is recommended that you validate the protocol experimentally. For example, the phalanx hybridization system, the MAUI System from BioMicro Systems, or HS Series of Hybridization Stations from TECAN offer a higher throughput and more automated hybridization methods.***

To complete this step, you will need to select a type of glass cover slide. Table 5, below, contains a list of glass cover slides that have been tested and confirmed compatible for use with the OneArray® Buffer.

Table 5: Compatible Glass Cover Slide Products

Manufacturer	Product Name
BioRad® Laboratories	SLS 6001 (24x60 mm)
Erie Scientific Company®	mSeries LifterSlip™ 25x601-M-5439
Corning®	Cover Glass (24 X 60 mm)

1) Ensure your work and experimentation area, as well as the OneArray®, are clean before adding the Hybridization Buffer solution to the target array.

2) Pre-warm the Hybridization Buffer with formamide at 42°C for 10 minutes.

3) Prepare the hybridization mix in a 1.5 ml Eppendorf tube according to the Table 6, below.

Table 6: Hybridization Mix Measurements	
For each slide: 55 µl	
Component	***Final Volume***
1.5X OneArray® Hybridization Buffer	**37 µl**
Sheared Salmon Sperm DNA (10 µg/µl)*	**1 µl**
Target preparation plus nuclease-free ddH_2O	**17 µl**

* Alternatives to Salmon Sperm DNA Blocking Mixtures:
Ambion® sheared Salmon Sperm DNA (10 µg/µl), or
Invitrogen™ Cot-1 DNA® (2.5 10 µg/µl), or
Invitrogen™ Poly-A (2.5 10 µg/µl)

4) Spin down the mixture for 5 minutes to eliminate potential debris.

5) Transfer the mixture to a new tube.

6) Heat the mixture to 95°C for 5 minutes (thermocycler recommended).

7) Maintain the mixture at a temperature of 60°C until pipetting onto the array (thermocycler recommended[1]).

[1] It may be helpful to set a Denature program in the thermocycler as follows:
95°C – 5 minutes
60°C – Hold

8) Place the OneArray® slide, bar code up, atop the "Probe Printed Region Guide" (included, see Figure 1).

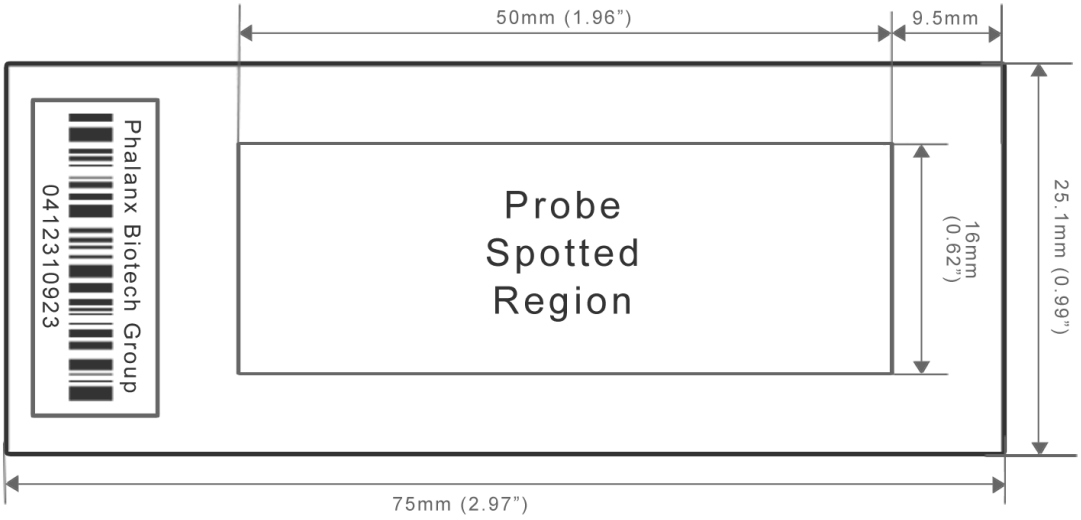


Figure 6: OneArray® Microarray Glass Slide with "Probe Printed Region Guide" Plastic Underlay.

9) Pipette the hybridization mixture onto the spotted region of OneArray® DNA Microarray. Avoid creating any bubbles.

10) Carefully place the glass cover slide over the spotted area in an even manner.

11) Place the entire labeled target plus the microarray set-up into a closable, chambered box* that is humidified by 2X SSPE buffer in the **50**℃ oven for 14 to 16 hours. A sealed chamber ensures that the appropriate humidity level is maintained during incubation. (See Figure 7).

Figure 7, below, provides an illustration of Step 4Bc, where the hybridization protocol is completed using the glass cover slide method, and specifically, the OneArray® DNA Microarray is placed into the chambered box.

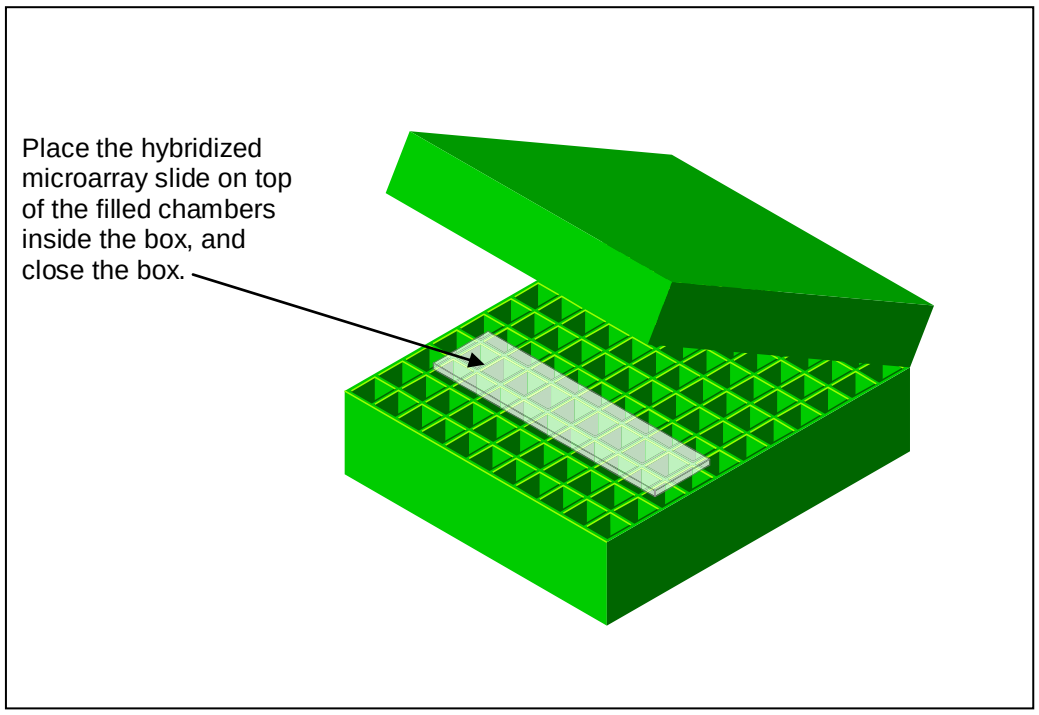


Figure 7: Step 4Cc→ aRNA Hybridization—Glass Slide Inside Chamber Box[2]

[2] The Hinged 100-Place Storage & Freezer Polypropylene Box from USA Scientific has been used to complete this step with frequent success. The small (approximately ½ inch x ½ inch) chambers within the box are filled about ¾ full of buffer, then the microarrays are laid on top of the chambers. The box is then closed and placed inside the oven. For information about this product or other USA Scientific products, access their Web site at: www.usasciesntific.com

D: Hybridization Using the MAUI hybridization system

Step 4Da: →Prepare Hybridization Solution Using the OneArray® Hybridization Buffer (Included)

For correct use of this buffer, you must add a specific amount of formamide and labeled target. Please follow the instructions below carefully.

1) Spin down the stock OneArray® Hybridization Buffer (~ 5.5 ml in each tube).

2) Add 4.5 ml of deionized formamide.

3) Warm the mixture to 42°C to completely dissolve the solution. Mix thoroughly.
Yield: 10 ml of 1.5X Hybridization Buffer solution.

4) Make up 1 X Hybridization Buffer by adding nuclease-free H_2O.

5) Aliquot the solution into individual tubes according to usage and store in darkness at -20°C.

Step 4Db: →Prepare Target for Hybridization

✧Hybridization Using Labeled Targets from aRNA Labeling Approaches

1) Mix 2 µg of your aRNA sample with nuclease-free H_2O to yield a final volume of 9 µL.

NOTE: ***It is essential to use at least 2 µg of labeled target for each hybridization.***

2) Add 1 µl 10x Fragmentation Reagent, and incubate at 70°C for 15 minutes.

3) Add 1 µl Stop Solution, and mix well.

4) Mix with nuclease-free H_2O to yield a final volume of 14.2 µL.

5) Keep on ice and in darkness until hybridization (Step 4Dc).

Step 4Dc: →Complete the Hybridization

1) Ensure your work and experimentation area, as well as the OneArray®, are clean before adding the Hybridization Buffer solution to the target array.

2) Pre-warm the Hybridization Buffer with formamide at 42°C for 10 minutes.

3) Prepare the hybridization mix in a 1.5 ml Eppendorf tube according to the Table 7, below.

Table 7: Hybridization Mix Measurements	
For each slide: 45 µl	
Component	***Final Volume***
1.5X OneArray® Hybridization Buffer	**30 µl**
Sheared Salmon Sperm DNA (10 µg/µl)*	**0.8 µl**
Target preparation plus nuclease-free ddH_2O	**14 .2 µl**

* Alternatives to Salmon Sperm DNA Blocking Mixtures:
Ambion® sheared Salmon Sperm DNA (10 µg/µl), or
Invitrogen™ Cot-1 DNA® (2.5 10 µg/µl), or
Invitrogen™ Poly-A (2.5 10 µg/µl)

4) Spin down the mixture for 5 minutes to eliminate potential debris.

5) Transfer the mixture to a new tube.

6) Heat the mixture to 95°C for 5 minutes (thermocycler recommended).

7) Maintain the mixture at a temperature of 60°C until pipetting onto the array (thermocycler recommended[3]).

8) Load the MAUI SC Mixer as described in the MAUI Hybridization System User's Guide (CDP-14).

9) After sealing the fill and vent ports with port seals, place the MAUI Mixer-Slide Assembly on a pre-warmed (50 °C) MAUI Hybridization System. Close the MAUI Hybridization System lid and hybridize for 4 – 12 hours (4-Bay = Mix Mode B, 12-Bay = Mix Mode A).

10) Following hybridization, disassemble the MAUI Mixer-Slide Assembly under pre-warmed (42°C) Wash Buffer #1

Step 5: Wash the Hybridized Microarray

Washed and dried microarrays should be scanned within a couple of hours.

NOTE: ***Do not allow the microarray(s) to be exposed to air for a significant amount of time; otherwise, an increased fluorescent background signal could appear.***

1) Submerge the entire labeled target and microarray set-up with the cover slide still intact into a large container filled with 42°C 2X SSC, 0.2% SDS solution.

2) Carefully remove the cover slide from the glass by gently shaking the glass slide so that the cover slide is freed while the slide is submerged.

NOTE*: At this stage, the microarray has the highest concentration of unhybridized target and dye. Transfer the array quickly to the slide rack to minimize exposure to air.*

3) Wash the slide(s) in the "rectangular, slide staining dish and slide rack" with the excess amount of pre-warmed 2X SSC, 0.2% SDS solution for 5 min at 42℃.

4) Transfer the slide rack to a second slide staining dish that contains 2X SSC wash for 5 min at 42℃.

5) Transfer the slide rack to a third slide staining dish that contains 2X SSC and wash for 5 min at room temperature.

6) Rinse each array carefully with 0.2X SSC using a squeeze bottle.

7) Spin dry with a centrifuge for at least one minute.

8) Keep the microarray dry and in the dark until ready to scan.

Step 6:

Scan and Extract Gene Expression Results

There are many scanners available to extract signals from OneArray®. Data extraction using GenePix™ 4100 from Molecular Devices is described below. Please refer to the respective company product instructions for appropriate use.

Table 8, below, lists the setting for using the GenePix 4100.

For a list of scanners that are compatible with the OneArray®, please refer to Table 8, below.

NOTE: *The performance of each scanner may differ. Therefore, to ensure best results, it is recommended that the scanner be adjusted based on standard microarray calibration parameters. Turn on and warm up the scanner for the duration according to manufacture instructions for the scanner.*

Use the .gal file and Gene List provided with this product, or refer to our website at:

www.onearray.com

Table 8: Scanner Settings Using GenePix™ 4100 from Molecular Devices

Wavelength	**635 nm**	**532 nm**
PMT	**630 V**	**590 V**
Minimum diameter (%)	**50**	
Maximum diameter (%)	**200**	
CPI Threshold	**0**	

NOTE: *For lower versions of GenePix software, adjust the property parameter to 142.8 μm manually to obtain best results.*

Figure 8, below provides a visual example of the OneArray® glass slide with spotted probe region.

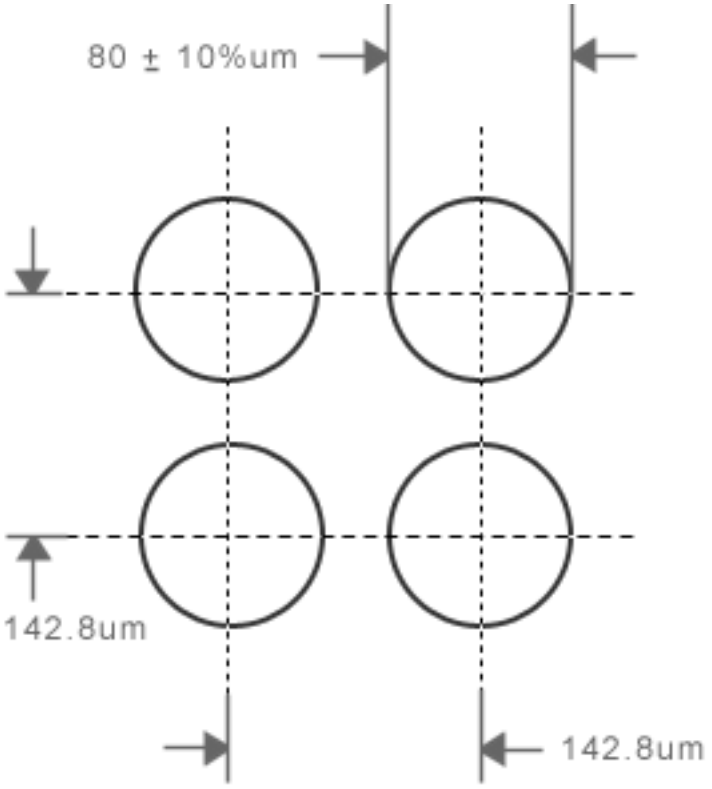


Figure 8: OneArray® Glass Slide with Spotted Probe Region.

OneArray® Microarray Scanner Specifications

Select and use a microarray scanner that meets the specifications below.

Microarray Scanner Specifications	
Format capabilities:	**1” x 3” (one inch by three inch) glass slide**
Molecular capabilities:	**Able to accurately detect, activate and read Cy3 and Cy5 fluorescent molecules**

Table 9, below, contains a partial list of microarray scanner products that are compatible for use with the OneArray® microarray. Please refer to the respective company website for more information about the products listed below.

Table 9: Compatible Microarray Scanners	
Manufacturer	**Product Name and Description**
Molecular Devices	**Axon GenePix® 4000, 4100, and 4200 series**
Genomic Solutions,® Inc.	**GeneTAC™ 2000**
Perkin Elmer,® Inc.	**ScanArray™ 5000**
TECAN®	**LS 200/300/400**
Agilent Technology	**DNA Microarray Scanner G2565B**

Step 7: Check the Control Probe Data

OneArray® DNA Microarrays contains built-in control probes for performance monitoring of the hybridization process. They are used to confirm or deny whether the experiment was completed successfully. Please visit

http://www.phalanx.com.tw/Support/RiOA_CP.php l

for more detailed information about the experimental controls on your OneArray® product.

Additional information about the control probes is included on the Product Support CD, and on our website at:

www.onearray.com

OneArray® Product Family

- **Human OneArray® v5**

  - 29,187 human genome probes
  - 1,088 experimental control probes
  - Composition: RefSeq release 38 and Ensembl release 56

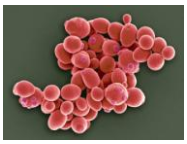
- **Mouse OneArray® v2**

  - 26,423 mouse genome probes
  - 872 experimental control probes
  - Composition: RefSeq release 42 and Ensembl release 59

- **Rat OneArray® v1**

  - 24,358 rat genome probes
  - 980 experimental control probes
  - Composition: RefSeq release 42 and Ensembl release 59

- **Yeast OneArray® v1**



  - 6,958 yeast genome probes
  - 684 experimental control probes
  - Composition: AROS v1.1 and YBOX v1.0

- **Rice OneArray® v1**

  - 22,003 rice genome probes
  - 824 experimental control probes
  - Composition: RGAP v6.1 and BGI 2008

- **Human miRNA OneArray® v2**

  - 1,087 unique miRNA probes
  - 105 experimental control probes
  - 3 features per probe
  - 100% of Sanger miRBase v15 Human miRNAs

- **Mouse & Rat miRNA OneArray® v2**

  - 785 unique miRNA probes
  - 105 experimental control probes
  - 3 features per probe
  - 100% of Sanger miRBase v15 miRNAs

Power of OneArray

OneArray®

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